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### *Escherichia coli* Antisera

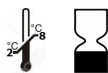
*E. coli* antisera are produced for serological identification of *E. coli*, based on agglutination method. These polyclonal antibodies are prepared by immunizing rabbit with the standard strains.

For high specificity; the non-specific agglutinins have been removed by absorption.

Preservative : 0.1% Sodium azide

Storage condition : 2-8°C / See Exp. Date on package

Do not freeze the antisera.



Complete identifications of *E. coli* requires cultural isolation, biochemical characterization and serotyping. However well defined the serology, the use of serological procedures do not supersede cultural isolation and biochemical characterization.

### Principle of the procedure

Serological tests are based on the fact that antibodies in serum, produced in response to exposure to bacterial antigens, will agglutinate with bacteria carrying homologous antigens. Test cultures with polyvalent sera, which are intended for use by the slide agglutination technique only. Both the confluent growth and selected colonies from the primary plate should be examined.

A positive slide reaction with a live culture may be due to the presence of K antigen on the surface of the organisms.

### O-antigen testing

#### A. Slide agglutination of live organisms

1. Place 1-2 drops of saline onto control area and place a drop of polyvalent antiserum onto test area of clean glass slide.
2. Using platinum wire, transfer a portion of a loopful of growth from TSI slant onto the drop of saline and antiserum, mix the cultures and serum or cultures and saline well. Tilt the glass slide back and forth for one minute. If there is clumping in the control, the culture is rough and serological tests cannot be interpreted on slide testing.
3. If the live organisms doesn't give positive reaction with antiserum, mix 2 loops of the culture with 1 ml. of saline then heat at 100°C for 1 hr. then repeat step 1 and 2.
4. Further testing of the isolate should be conducted as described in steps 1-3 with monovalent O antisera to reveal the full O antigenic grouping of the isolate. Always confirm the O grouping by slide agglutination on heat killed organisms, see next step.

#### B. Slide agglutination of heat killed organisms

If the live organisms give positive reaction with antiserum, mix 2 loops of the culture with 1 ml. of saline then heat at 100°C for 1hr. then repeat the agglutination test as above using monovalent O antiserum on the heated cell suspension. This should be done to identify the O antigen type as distinct from the K antigen.

### Slide Agglutination Interpretation

Agglutination should be strong and clearly visible within 1 minute. There should be no visible agglutination in the control suspension ; if agglutination is seen in the control, the suspension is not suitable for testing by this method.

### H-antigen Testing

1. Allow the organism to pass through semi-solid medium. Then inoculate the resulting organism into 2 ml of Tryptic soy broth tubes and incubate at 37°C for 6-8 hrs.
2. After incubation period, add 2 ml. of 0.85% saline containing 1% (v/v) Formalin, allow the inoculum at the room temperature for 30 minutes. Transfer the 0.45-0.5 ml of inoculum into duplicate of 12x75 mm tube.
3. For the first tube, add 2 drops of *E. coli* H7 antiserum (AS546) and not add any antiserum for another tube for Negative control. Allow both tube to stand in water bath at 48-50 °C for 1 hour.
4. After incubation period, observe the result by checking the negative control tube which should shown an even suspension; clumping in the control indicates that the culture is rough and serological tests cannot be interpreted, then observe another tube for the flocculation reaction; that is Positive result. An isolate producing a distinct positive reaction is assumed to be an *E. coli* bearing the H antigenic factors represented by that antiserum.

### *E. coli* Polyvalent Antisera

Cat. No.	Description	Specific Factors
AS521	<i>E. coli</i> (O&K) Polyvalent I	O25 : K11
		O26 : K60
		O44 : K74
		O55 : K59
		O78 : K80
		O111 : K58
		O114 : K-
AS530	<i>E. coli</i> (O&K) Polyvalent II	O119 : K69
		O86 : K61
		O124 : K72
		O125 : K70
		O126 : K71
		O127 : K63
		O128 : K67
AS537	<i>E. coli</i> (O&K) Polyvalent III	O18a O18c : K77
		O20a O20b : K84
		O28 : K73
		O112a O112c : K66

### *E. coli* Monovalent Antisera

Cat. No.	Description	Cat. No.	Description
AS522	<i>E. coli</i> O25 : K11	AS534	<i>E. coli</i> O126 : K71
AS523	<i>E. coli</i> O26 : K60	AS535	<i>E. coli</i> O127 : K63
AS524	<i>E. coli</i> O44 : K74	AS536	<i>E. coli</i> O128 : K67
AS525	<i>E. coli</i> O55 : K59	AS538	<i>E. coli</i> O18a O18c : K77
AS526	<i>E. coli</i> O78 : K80	AS539	<i>E. coli</i> O20a O20b : K84
AS527	<i>E. coli</i> O111 : K58	AS540	<i>E. coli</i> O28 : K73
AS528	<i>E. coli</i> O114 : K-	AS541	<i>E. coli</i> O112a O112c : K66
AS529	<i>E. coli</i> O119 : K69	AS545	<i>E. coli</i> O157
AS531	<i>E. coli</i> O86 : K61	AS546	<i>E. coli</i> H7
AS532	<i>E. coli</i> O124 : K72	AS547	<i>E. coli</i> O1 : K1
AS533	<i>E. coli</i> O125 : K70	AS548	<i>E. coli</i> O2

### Reference:

1. Kauffmann, F., Classification of Bacteria 1947
2. Edward, P.R. and Ewing, W.H., Identification of Enterobacteriaceae, Fourth Edition, Burgess Company, Minnesota 1986

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